A Summary of Fungal Leaf Pathogens of *Eucalyptus* and the Diseases they Cause in South Africa*

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Eucalyptus leaf disease surveys conducted since 1984 in the Cape, Eastern Transvaal and Natal Forest Regions (Crous, Knox-Davies and Wingfield. 1988, 1989a, b, c) revealed a number of fungal leaf pathogens previously unreported in South Africa. This paper combines information gained during these surveys with records from previous sources (Doidge, 1950; Doidge et al., 1953; Lundquist and Baxter, 1985). We also include a key to these Eucalyptus leaf pathogens. Opportunistic fungi such as Botryosphaeria ribis Grossenb. & Duggar, Botrytis cinerea Pers., Cylindrocladium spp. and Hainesia lythri (Desm.) Höhnel., which are commonly found on Eucalyptus leaves under stress conditions and which have a wide host range, have not been included.

Where possible, comment is made on the relative significance of the pathogens to *Eucalyptus* propagation. These evaluations are based on field observations, the relative importance of the *Eucalyptus* spp. affected, and reports from other countries.

Aulographina eucalypti (Cooke & Massee) von Arx & Muller

Anamorph: Thyrinula eucalypti (Cooke & Massee) Swart.

Occurrence. Most common in the Transvaal and Natal; not reported further south than the Eastern Cape.

Symptoms. Causes a severe leaf spotting of mature leaves (Figure 1). Lesions seldom penetrate through the leaf lamina and are distinct, brown, circular and corky. A. eucalypti often occurs in association with Mycosphaerella nubilosa (Cke.) Hansf.

Pathogen. Amphigenous hysterothecia and pycnidia occur on older lesions (Figure 2). Ascospores are hyaline, 2-celled, constricted at the septum, rounded at both ends and measure 9-(13)-16x3,5-(4)-5 μ m (Figure 3).

Hosts. E. andrewsii Maid., E. coriaceae A.Cunn., E. dalrympleana Maid., E. elata Dehnh., E. fastigata Deane & Maid., E. gigantea Hooke f., E. globoidea Blakely, E. globulus Labill., E. hemiphloia F. Muell., E. macarthurii Deane & Maid., E. muelleriana Howitt, E. nitens (Deane & Maid.) Maid., E. obligua L. Heritier, E. oreades R.T. Bak., E. pilularis Sm., E. quadrangulata Deane & Maid., E. regnans F. Muell., E. resinifera Sm., E. saligna Sm.

Relative importance. Causes extensive defolation, and can thus be of economic importance.

Pseudocercospora eucalyptorum Crous, Wingfield, Marasas & Sutton

Occurrence. Widely distributed throughout the Cape

Province and Natal. Infects a number of *Eucalyptus* spp., although it occurs most commonly on the older leaves of *E. nitens* (Crous, Wingfield, Marasas & Sutton, 1989).

Symptoms. Symptoms vary on the different hosts. Leaf spots range from subcircular and discrete to confluent, but on *E. nitens* they are always angular and confined by the leaf veins (Figure 4). Variable in colour from chlorotic to light brown and grey-brown, depending on age.

Pathogen. Grey to brown tufts of conidiophores are found on older lesions (Figure 5). Conidia are olivaceous, cylindrical, straight to slightly curved, indistinctly 1-6 septate (Figure 6), measure $23-(42)-65x2,5-(3,5)-4\mu m$.

Hosts. E. bridgesiana R.T. Bak., E. deanei Maid., E. nova-anglica Deane & Maid., E. nitens, E. pellita F. Muell.

Relative importance. At present P. eucalyptorum does not seem to be of any economic importance.

Coniothyrium ovatum Swart

Occurrence. As yet, only found in the Western and Southern Cape.

Symptoms. Occurs on immature leaves on young growth and the lower branches of mature trees. Lesions are irregular, dark purple to black on the upper leaf surface, and light to dark brown on the lower surface (Figure 7).

Pathogen. Of the five Coniothyrium spp. occurring on Eucalyptus (Sutton, 1971, 1975; Swart, 1986), only C. ovatum has been reported from South Africa (Crous, Knox-Davies and Wingfield, 1988). Substomatal pycnidia, which are prominent on the lower leaf surface, exude long black cirri of slightly roughened, dark brown conidia (Figure 8). Conidia are obovate with truncate bases and measure 7-(8,5)-10x5-(5,2)- 6μ m (Figure 9).

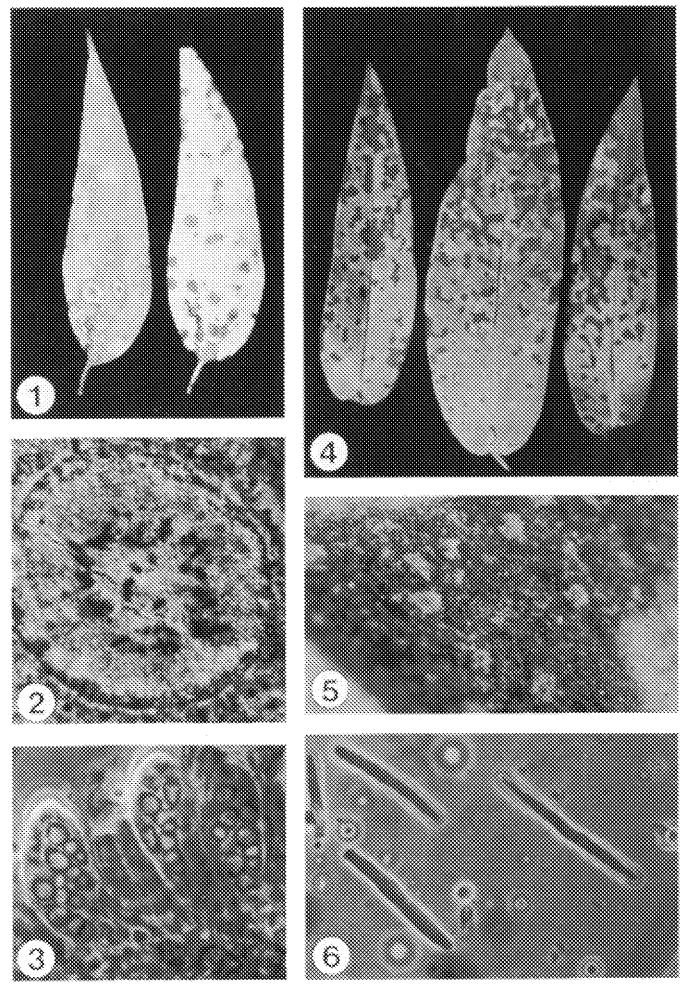
Hosts. E. cladocalyx, E. lehmannii Preiss: Schauer.

Relative importance. It seems likely that trees will outgrow this pathogen owing to its inability to infect actively-growing mature foliage. The relative unim-

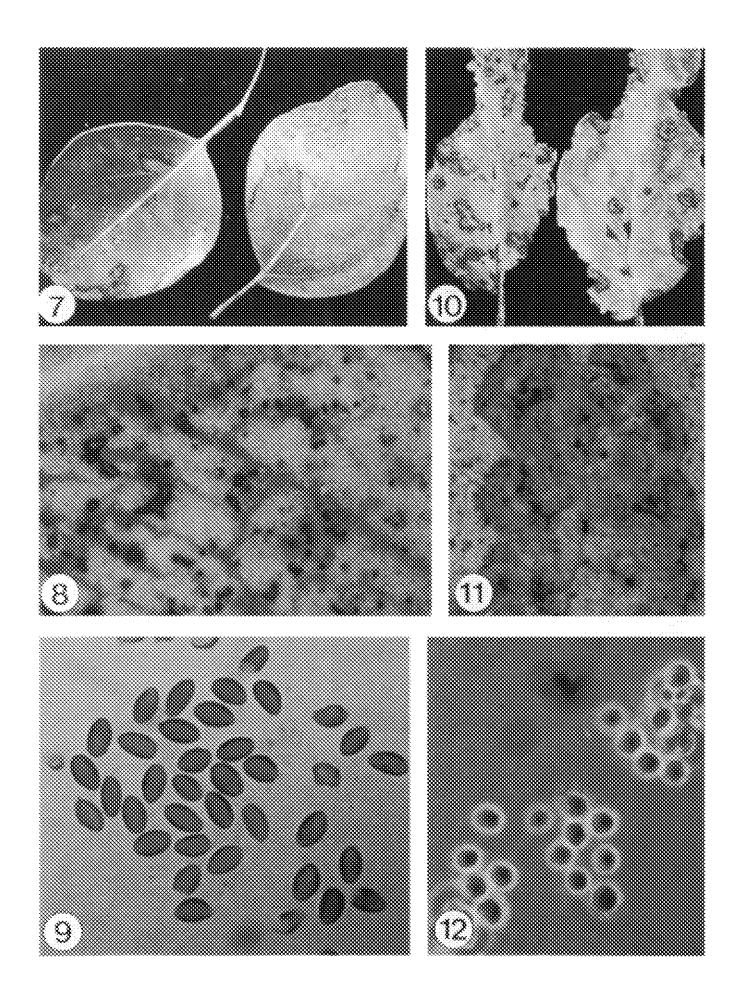
^{*} Part of an M.Sc. Agric. thesis submitted by the first author to the University of Stellenbosch.

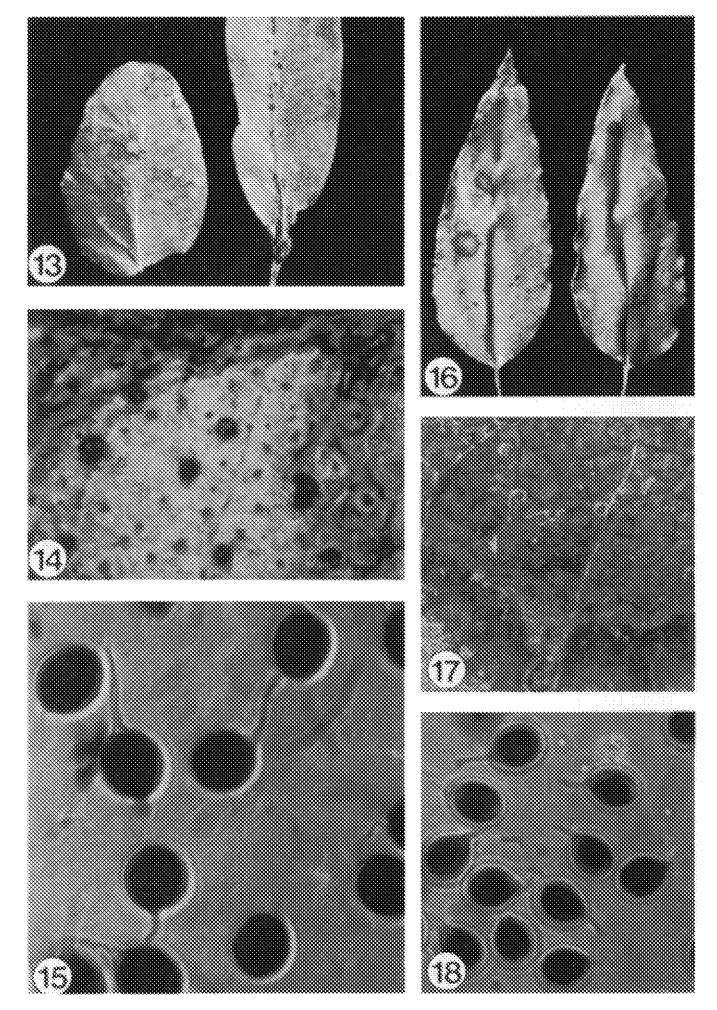
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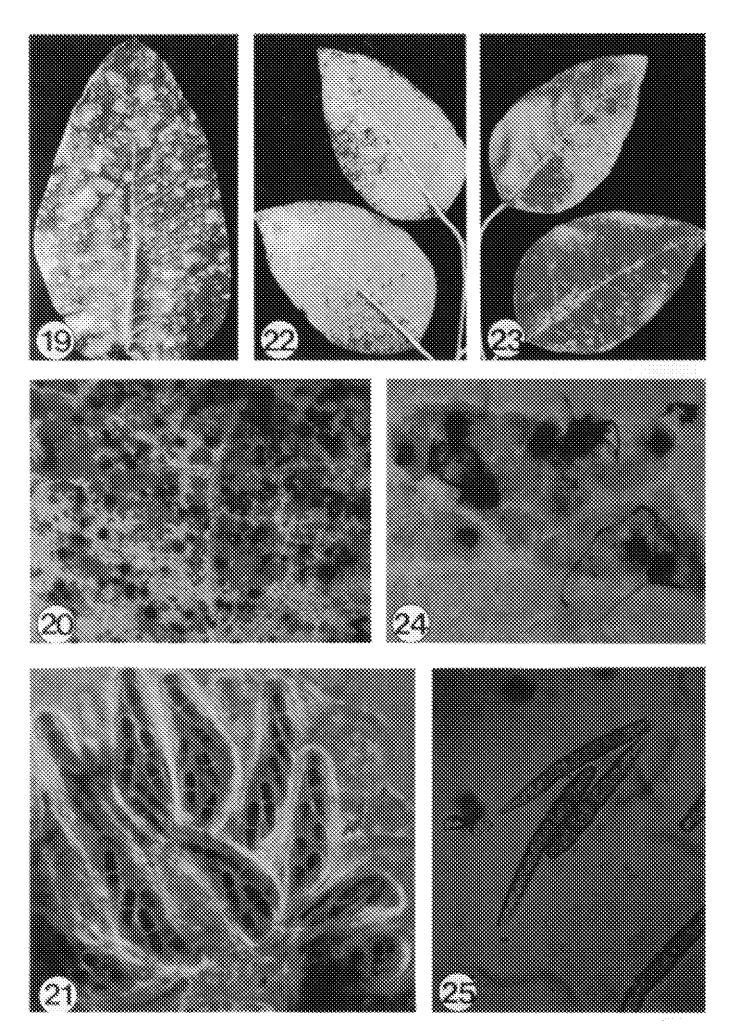
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- FIGURE 1. Symptoms caused by Aulographina euclypti on young and older leaves of E. nitens.
- FIGURE 2. Hysterothecia and pycnidia of A. eucalvpti (x360).
- FIGURE 3. Asci and ascospores of A. eucalypti (x2250).
- FIGURE 4. Angular lesions caused by Pseudocercospora eucalyptorum on older leaves of E. nitens
- FIGURE 5. Grey-brown tufts of conidiophores of P. eucalyptorum (x720).
- FIGURE 6. Conidia of P. eucalyptorum (x2 250)
- FIGURE 7. Symptoms caused by Coniothyrium ovatum on E. cladocalyx.
- FIGURE 8. Exuding conidia of C. ovatum (x560).
- FIGURE 9. Conidia of C. ovatum (x2250).
- FIGURE 10. Symptoms caused by Fairmaniella leprosa on leaves of E. globulus.
- FIGURE 11. Acervuli of F. leprosa (x175).
- FIGURE 12. Conidia of F. leprosa (x2 250).
- FIGURE 13: Symptoms caused by Harknessia eucalypti on leaves of E. maidenii (left) and E. nitens (right).

- FIGURE 14. Fructifications of H. eucalypti (x500).
- FIGURE 15. Conidia of H. eucalypti (x2 250).
- FIGURE 16. Symptoms caused by Harknessia globosa on leaves of E. nitens clone.
- FIGURE 17. Fructifications of H. globosa (x300).
- FIGURE 18. Conidia of H. globosa (x2 250).
- FIGURE 19. Symptoms caused by Mycosphaerella nubilosa on E. nitens.
- FIGURE 20. Pseudothecia of M. nubilosa (x560).
- FIGURE 21. Asci and ascospores of M. nubilosa (x2 250).
- FIGURE 22. Symptoms caused by Phaeoseptoria eucalypti on the lower surface of E. grandis leaves.
- FIGURE 23. Symptoms caused by P. eucalypti on the upper surface of E. grandis leaves.
- FIGURE 24. Exuding conidia of P. eucalypti (x720).
- FIGURE 25. Conidia of P. eucalypti (x1 750).
- FIGURE 26. Leaves covered with white mycelium of Sphaerotheca pannosa.
- FIGURE 27. Conidia of S. pannosa (x1 750).

portance of recorded hosts also reduces its potential significance.

Fairmaniella leprosa (Fairm.) Petrak & Syd.

Occurrence. At present only known from Franschhoek and Stellenbosch in the Cape Province (Crous, Knox-Davies and Wingfield, 1989c). Here it occurs on mature, older leaves 4 m above the ground, below the actively-growing crown.

Symptoms. Hard, round, corky, brown lesions on the leaf lamina and petioles (Figure 10). Lesions are distinct and usually do not extend through the lamina.

Pathogen. Amphigenous acervuli occur on lesions (Figure 11), with brown, thick-walled conidia varying in shape from elongate to broadly elliptical. Conidia measure $4-(5)-6.5\times3-(4)-4.5 \mu m$ (Figure 12).

Relative importance. Judging from the limited host range and distribution, this fungus appears to be a highly specific pathogen. It is, therefore, doubtful if it will be of great economic importance in South Africa.

Harknessia eucalypti Cke. apud Cke. & Hark.

Occurrence. On mature and juvenile leaves of Eucalyptus spp. at Stellenbosch (Crous, Knox-Davies and Wingfield, 1989c).

Symptoms. Prominent leaf and stem necrosis; lesions distinct, light brown, round to irregular in shape, penetrating through the lamina (Figure 13).

Pathogen. Fructifications extrude black conidial masses (Figure 14). Conidia dark brown, smoothwalled, broadly ventricose with bluntly apiculate apices, measuring $16-(19)-22x8-(12)-14\mu m$. Hyaline appendages 2-(8.5)-18 μm long (Figure 15). Unlike many other Eucalyptus leaf pathogens. H. eucalypti can grow in culture.

Hosts. E. globulus, E. maidenii F. Muell., E. nitens. Relative importance. Unknown.

Harknessia globosa Sutton

Occurrence. Only found in nurseries in the Eastern Transvaal (Crous, Knox-Davies and Wingfield, 1989c).

Symptoms. Prominent leaf spots on young *E. grandis* Hill: Maid.; Lesions round, amphigenous, brown and 5-15 mm in diameter (Figure 16).

Pathogen. Fructifications occur on older lesions, extruding black conidial masses (Figure 17). Conidia dark brown, smooth-walled, globose to subglobose, measuring $10-(13,5)-15x9-(11)-13\mu m$. Hyaline appendages are $1-(4)-9 \mu m \log (Figure 18)$.

Hosts. E. grandis

Relative importance. Only found in nurseries, which suggests that it is of minor importance.

Mycosphaerella nubilosa (Cke.) Hansf.

Occurrence. Appears to be the only Mycosphaerella

sp. occurring on *Eucalyptus* spp. in South Africa (Crous, Knox-Davies and Wingfield, 1989b) and is found throughout the country.

Symptoms. Occurs mainly on immature foliage, where it causes large yellow to brown lesions and a twisting of the leaf lamina (Park and Keane, 1982). In South Africa, lesions vary from pin-head spots to round or irregular spots, which coalesce to form large blotches (Figure 19).

Pathogen. Pseudothecia occur on both the upper and lower surfaces of the lesions (Figure 20). Asci measure 35-(54)-63x8-(10)-15 μ m. Ascospores not constricted at the septum, tapering towards the base and measuring 9-(13,5)-18x3-(3,5)-4 μ m (Figure 21).

Hosts. E. bicostata Maid. et al., E. camaldulensis Dehnh., E. dalrympleana, E. globulus, E. grandis, E. grandis x nitens hybrids, E. macarthurii, E. maidenii, E. nitens, E. nova-anglica, E. saligna, E. smithii R.T. Bak., E. stuartiana F. Muell., E. tereticornis Sm., E. viminalis Labill.

Relative importance. Young E. nitens (Deane & Maid.) Maid. trees of the Victoria Provenances are highly susceptible to infection which usually leads to total loss of juvenile leaves (Lundquist and Purnell, 1987). Mature leaves can also be infected. M. nubilosa has severely affected the propagation of E. nitens and E. globulus in South Africa. Its common occurrence on E. grandis and hybrids of E. grandis and E. nitens is of concern.

Phaeoseptoria eucalypti Hansf. emend. Walker

Occurrence. Found on Eucalyptus spp. throughout South Africa (Crous, Knox-Davies and Wingfield, 1988). Usually occurs only on the lower leaves, below the actively growing crown.

Symptoms. Lesions irregular, brown, and surrounded by a prominent purple discolouration (Figure 22, 23).

Pathogen. Substomatal pycnidia, which are prominent on the lower leaf lamina, exude long black cirri of sligtly roughened conidia (Figure 24). Conidia elongate with a rounded to attenuate apex and subtruncate base, 40-(47)-55x4-(5)-6 μ m, with 3-(4)-7 transverse septa (Figure 25).

Hosts. E. bicostata, E. camaldulensis, E. cladocalyx, E. dunnii Maid., E. globulus, E. grandis, E. grandis x camaldulensis, E. grandis x cladocalyx, E. grandis x nitens, E. grandis x tereticornis, E. grandis x urophylla, E. macarthurii, E. macarthurii x grandis, E. macarthurii x tereticornis, E. maidenii, E. nitens, E. nova-anglica, E. quadrangulata, E. resinifera, E. saligna, E. tereticornis.

Relative importance. Causes a severe infection of younger leaves of some E. grandis clones. It can also cause severe damage to seedlings in nurseries (Sharma, Mohanan and Florence, 1984) and hedges used in clonal propagation (Crous, Knox-Davies and Wingfield), 1988). It remains doubtful whether P. eucalypti can completely defoliate healthy mature trees, but there is

evidence to suggest that it becomes more important when trees are under stress. Because of the significance of *E. grandis* to the South African forestry industry, this fungus should be considered one of the more important *Eucalyptus* leaf pathogens.

Sphaerotheca pannosa (Wallr.:Fr.) Lev.

Occurrence. Although powdery mildew is well known on *Eucalyptus* spp. (Boesewinkel, 1981), it has only recently been reported from South Africa, (Crous, Knox-Davies and Wingfield, 1989a).

Symptoms. Leaves are covered with a densely interwoven white mycelium. The fungus favours young leaves and shoots (Figure 26); however, Boesewinkel (1981) and Gibson (1975) report that it can cause spotting and malformation of older growth.

Pathogen. Conidia produced in chains (4-8 per chain), and ellipsoidal, hyaline, measuring 21-(29)-35x12-(15)-19 μ m (Figure 27).

Hosts. E. nitens, E. nitens x camaldulensis, Eucalyptus sp.

Relative importance. S. pannosa (rose mildew) occurs naturally on roses, but has adapted to infect eucalypts. This fungus has been recorded only once, and its importance is unknown. G.J.M.A. Gorter (Plant Protection Research Institute, Pretoria, personal communication) is of the opinion that it is the same "form" which usually infects peaches.

KEY TO EUCALYPTUS LEAF PATHOGENS IN SOUTH AFRICA

1. Leaves covered with densely interwoven white my-

celium, which turns brown with age; conidia in

	chains, ellipsoidal, hyaline
	(Fig. 27) Sphaerotheca pannosa
	Not as above, fungus causing leaf and stem necro-
	sis
2.	Spores predominantly conidia
	Spores predominantly ascospores
3.	Conidia single-celled4
	Conidia septate
4.	Conidia with appendages
5.	Conidia without appendages9
6.	Conidia, cylindrical, straight to slightly curved, in-
7.	distinctly 1-6 septate; borne on amphigenous tufts of conidiophores on older lesions, mostly greyish (Fig. 6)

	(Fig. 21) Mycosphaerella nubilosa
8.	Conidia dark brown, smooth-walled, broadly ven-
	tricose with bluntly apiculate apices and hyaline
	appendages 2-(8,5)-18 m
	(Fig. 15) Harknessia eucalypti
	Conidia dark brown, smooth-walled, globose to
	subglobose with hyaline appendages $1-(4)-9\mu m$
	(Fig. 18) Harknessia globosa
9.	Pycnidia hypophyllous, substomatal, exuding long
	black cirri of slightly roughened obovate conidia
	with truncate bases
	(Fig. 9) Coniothyrium ovatum
	Acervuli amphigenous; conidia brown, thick-
	walled, elongate to broadly ellipsoidal
	(Fig. 12) Fairmaniella leprosa
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