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Departments of Plant Pathology and Microbiology and Biochemistry, University of the Orange Free State, Bloemfontein, South Africa

# Cryphonectria cubensis, a potential pathogen of Psidium guajava in South Africa

By W. J. Swart, E. Conradie and M. J. Wingfield

### Abstract

The pathogenicity of an isolate of *Cryphonectria cubensis* was tested in artificial inoculations on *Eucalyptus grandis, Psidium guajava* and *Syzygium cordatum.* The isolate was able to colonize host tissue of all three species following inoculations of branch sections. It rapidly colonized the tissue of potted guava plants of the important commercial cultivar, Fan Retief and resulted in the death of a number of inoculated branches. Pathogenicity was also proven on mature guava trees in the field. An important implication of these findings is that the cultivation of guavas in South Africa could be confronted with a potentially severe disease problem in the future.

# 1 Introduction

*Cryphonectria cubensis* (Bruner) Hodges is an important pathogen of *Eucalyptus* spp. in many countries throughout the world (BRUNER 1916; BOERBOOM and MAAS 1970; HODGES et al. 1979; HODGES 1980; GIBSON 1981; DAVIDSON and TAY 1983; SHARMA et al. 1985 a and b; FLORENCE et al. 1986; HODGES et al. 1986; OLD et al. 1986; WINGFIELD et al. 1989). *C. cubensis* is also reported to occur naturally on myrtaceous hosts other than *Eucalyptus* spp. (HODGES et al. 1986; HODGES 1980). *Endothia eugeniae* (Nutman and Roberts) Reid and Booth, conspecific to *C. cubensis* (ALFENAS et al. 1984; HODGES et al. 1986; MICALES and STIPES 1984), has been associated with dieback of clove [Syzygium aromaticum (L.) Merr. and Perry] in Brazil and Indonesia (HODGES et al. 1986).

Since C. cubensis was recently discovered in South Africa (WINGFIELD et al. 1989), there has been speculation that the pathogen could spread to, or have originated, on other species of *Myrtaceae* in this country. HODGES et al. (1986) found that C. cubensis could infect numerous species of Syzygium following artificial inoculations. Three genera of indigenous Myrtaceae occur in South Africa viz., Eugenia L., Syzygium Gaertn., and Metrosideros Banks ex Gaertn. of which Syzygium is the most prominent and represented by Syzygium guineense (Willd.) DC. Exotic species such as guava (Psidium guajava L.) are planted commercially in S. A. on a large scale (PALMER and PITMAN 1973). The total guava production for this country is approximately 35000 tons per year of which the cultivar, Fan Retief delivers 98% of total crop (BOLT 1984b; DU PREEZ 1986). The ability of C. cubensis to infect myrtaceous hosts other than Eucalyptus spp. could thus pose a potential threat to guava cultivation in this country.

The main purpose of this study was to establish whether local isolates of *C. cubensis* could become a threat to other exotic or indigenous myrtaceous hosts in this country. The relative virulence of a local eucalyptus isolate of *C. cubensis* was therefore tested on *E. grandis*, *P. guajava* and *S. cordatum*.

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### 2 Methods

### 2.1 Inoculation of branch sections

Freshly cut branch sections (2–4 cm in diameter and 15 cm in length) of *P. guajava, E. grandis* and *S. cordatum* were artificially inoculated under laboratory conditions with a single-spore isolate of *C. cubensis* obtained from diseased *E. grandis* trees in Natal, South Africa. Prior to inoculation, sections were surface-disinfected by swabbing with 0,5% NaOCl and the ends coated with melted paraffin-wax. Each section was inoculated by removing a 10mm diameter cambial disc with a cork borer and placing a disc of equivalent size from a 5-day-old Potato-dextrose agar (PDA) culture of *C. cubensis* in the wound. Sterile PDA discs were placed in wounds which served as controls. Each inoculated wound was covered with masking-tape. Inoculated branch sections were incubated at 25°C and the length of the cambial discolouration was measured after 3 weeks. Isolations were made from the advancing edge of each lesion to verify the presence of *C. cubensis*. Ten replicates of each treatment were made. A two-way analysis of variance was performed on the data and Tukey's HSD procedure was used to separate means.

### 2.2 Glasshouse inoculations

Inoculum of *C. cubensis* was prepared by culturing the fungus in petri dishes on PDA overlaid with sterile gauze strips ( $10 \times 50$  mm). Twenty 2-year-old guava plants grown in pots under greenhouse conditions were wounded by lightly scraping off a length of bark ( $3 \times 10$  mm) from the stem of each plant. Fifteen plants were then inoculated by wrapping gauze colonized by *C. cubensis* around each wound. Sterile gauze strips were use for control plants. Parafilm (American National Can) was then wrapped around all gauze-covered wounds. After 6 weaks the gauze strips and surrounding bark were removed and the length of cambial discolouration measured. Isolations were made from discoloured tissue to verify the presence of *C. cubensis*. The experiment was conducted twice and a two factorial variance analysis was performed on the data of the two trials.

### 2.3 Field inoculation

Five-year-old guava trees were wounded 1.5 m above soil level, by removing a cambial disc with a 12 mm cork borer from each of two lateral branches having the same diameter. A 12 mm disc from a 5-year-old PDA culture of *C. cubensis* was placed in one wound and a sterile PDA disc in the other. All wounds were covered with masking-tape to prevent contamination and the inoculum from drying out. After 6 weeks the masking-tape and bark surrounding wounds was removed and the length of cambial discolouration measured. Isolations were made from the advancing edge of each cambial lesion to verify the presence of *C. cubensis*.

## 3 Results

### 3.1 Inoculation of branch sections

Cambial lesions were significantly shorter (P < 0.05) on *Syzygium* than on *Psidium* and *Eucalyptus* (Table 1, Fig. 1). Pycnidia had formed on the bark surrounding lesions of all three species. The mean length of cambial discolouration associated with the three species was significantly greater (P < 0.05) than that associated with controls (Fig. 2). Isolations from all lesions resulting from inoculation with *C. cubensis* revealed the presence of the pathogen.

Table 1. Lesion lengths on excised stems of three species of Myrtaceae after inoculation with Cryphonectria cubensis

Species	Lesion length (mm) <sup>1</sup>	
	Inoculation	Control
Psidium guajava	87.16 <sup>c</sup>	10.41ª
Eucalyptus grandis Syzygium cordatum	71.92°	11.66ª
Syzygium cordatum	44.50 <sup>b</sup>	10.06ª

### 3.2 Glasshouse inoculations

Six weeks after inoculation with *C. cubensis*, the mean length of cambial discolouration was significantly greater (P < 0.01) than that for the controls (Table 2). The foliage above inoculation points of five plants had died due to ring-girdling (Fig. 3) and pycnidia were visible on the bark surrounding the inoculated lesions. The pathogen was isolated from all lesions that had resulted from inoculation with *C. cubensis*.

# Table 2. Lesion lengths on stems of Psidium guajava after field and glasshouse inoculations with Cryphonectria cubensis

	Lesion length (mm) <sup>1</sup>	
	Inoculation	Control
Glasshouse inoculation		
Trial 1.	32.66ª	10 <sup>b</sup>
Trial 2.	29.17ª	10 <sup>b</sup>
Field inoculation	88.04ª	12 <sup>b</sup>

<sup>1</sup> Values are means of fifteen replicates. Values followed by different letters between columns are significantly different at P < 0.01. Glasshouse inoculation S. E. = 14.82; Field inoculation S. E. = 9.63

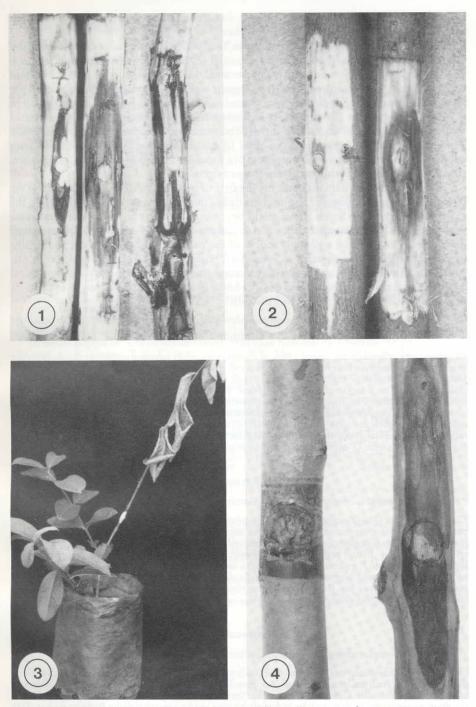
## 3.3 Field inoculations

All branches inoculated with *C. cubensis* displayed cambial lesions (Fig. 4), and pycnidia were observed on the surrounding bark. Control plants showed no sign of cambial discolouration and callus formation was far more advanced than in plants inoculated with *C. cubensis* (Table 2). The pathogen was re-isolated from all lesions that had resulted from inoculation with *C. cubensis*.

### 4 Discussion

The present study demonstrated that an isolate of *C. cubensis*, from cankers on *E. grandis* was able to colonize host tissue of *S. cordatum* and *P. guajava* following artificial inoculations of branch sections. HODGES et al. (1986) tested the pathogenicity of an isolate of *C. cubensis* originating from *E. saligna* Sm. in Hawaii on branch sections of 13 species of Myrtaceae including *P. guajava*. The pathogen grew on only four of species tested, excluding *P. guajava*. The disparity in the results of the two studies can probably be ascribed to differences between the isolate or cultivar used in each case.

It is significant that the isolate of *C. cubensis* used in the present study was able to cause lesions of approximately equivalent length on excised branches of mature *E. grandis* and *P. guajava* plants. The isolate, moreover, was able to colonize the tissue of potted guava



Figs. 1-4. Inoculation of three Myrtaceae genera with Cryphonectria cubensis. - Fig. 1. Lesions caused by C. cubensis on Syzygium, Eucalyptus and Psidium, from left to right. - Fig. 2. Control inoculation (l) and inoculation with C. cubensis (r) on branch section. - Fig. 3. Death of foliage above inoculation point 6 weeks after inoculation. - Fig. 4. Cambial lesion on branches of P. guajava following field inoculation (r) and control (l)

plants of the important commercial cultivar, Fan Retief relatively rapidly and even result in the death of a number of inoculated branches.

The potential for damage of *P. guajava* in South Africa by *C. cubensis* is noteworthy. Although the disease is favoured by high rainfall (2000–2400 mm/annum) and temperatures above 23 °C in other countries (FLORENCE et al. 1986; SHARMA et al. 1985 a) infection of *E. grandis* in S. A. commonly occurs in regions receiving less than 2000 mm rainfall per annum. In this country, *P. guajava* is cultivated in warm regions having a mean rainfall of 1000–2000 mm/annum (BOLT 1984 a). Furthermore, *C. cubensis* is frequently associated with natural or artificial wounds on clove roots, stems and branches (HODGES et al. 1986). Pruning of *P. guajava* is performed in order to modify the shape of trees, to boost fruit production and for regeneration purposes (BOLT and ALBERTS 1984). Pruning wounds are, therefore, potential infection courts for the pathogen.

The data presented here, indicate for the first time that *P. guajava* and *S. cordatum* are potential hosts for *C. cubensis*. The implications of these findings are threefold. Firstly, the cultivation of guavas in South Africa might be confronted with a potentially severe disease problem in the future; secondly, they fuel speculation (CONRADIE et al. 1989) that indigenous *Myrtaceae* could be a possible source of the pathogen in South Africa, and finally, indigenous *Myrtaceae* may serve as secondary hosts for *C. cubensis* from where it can spread to both *Eucalyptus* and *P. guajava* plantings. A survey to determine the geographical and ecological distribution patterns of *C. cubensis* in South Africa and thus to evaluate its potential impact must, therefore, be a priority.

### Summary

Cryphonectria cubensis is the causal agent of Eucalyptus canker in many countries throughout the world. The pathogen is also reported to infect other species of the Myrtaceae. This could pose a potential threat to exotic myrtaceous species such as guava (Psidium guajava) which are commercially cultivated on a large scale in South Africa. The main purpose of this study was, therefore, to establish whether a local, commercially important P. guajava cultivar and a common, indigenous member of the Myrtaceae, Syzygium cordatum were susceptible to C. cubensis. Artificial inoculation of branch sections of P. guajava and Syzygium cordatum with C. cubensis, resulted in rapid tissue colonisation by the pathogen. Results of artificial inoculations on young and mature P. guajava plants with C. cubensis, confirmed pathogenicity.

#### Résumé

# Cryphonectria cubensis, pathogène potentiel pour Psidium guajava en Afrique du Sud

*C. cubensis* est un agent de chancre chez les *Eucalyptus* dans de nombreuses régions du monde. Le pathogène est aussi mentionné chez d'autres espèces de *Myrtaceae*. Ceci peut constituer une menace pour certaines espèces exotiques telle que *Psidium guajava* qui est cultivé à grande échelle en Afrique du Sud. Le but principal de cette étude a été de savoir si un cultivar local, commercialement important de *P. guajava* et une Myrtacea indigène commune, *Syzygium cordatum*, étaient sensibles à *C. cubensis*. L'inoculation artificielle de sections de branches a conduit à une rapide colonisation des tissus par le pathogène. Les résultats d'inoculations effectuées sur des plants jeunes et matures de *P. guajava* ont confirmé le pouvoir pathogène de *C. cubensis*.

### Zusammenfassung

### Cryphonectria cubensis, ein potentielles Pathogen von Psidium guajava in Südafrika

Cryphonectria cubensis ist der Erreger von Eucalyptus-Krebs in vielen Ländern. Das Pathogen ist dafür bekannt, auch andere Arten aus der Familie der *Myrtaceae* zu befallen. Diese Tatsache könnte eine potentielle Gefahr für exotische Vertreter dieser Familie wie zum Beispiel Guava (*Psidium guajava*) darstellen, welche in Südafrika eine große wirtschaftliche Bedeutung hat. Das Hauptziel der vorliegenden Arbeit war es daher, festzustellen, ob eine lokal wirtschaftlich wichtige Sorte von *P. guajava* und *Syzygium cordatum*, ein weitverbreiteter einheimischer Vertreter der *Myrtaceae*, auf C. *cubensis* anfällig sind. Künstliche Inokulationen von Zweigabschnitten von P. guajava und S. cordatum resultierten in einer raschen Besiedlung durch den pathogenen Pilz. Die Resultate von Inokulationen von jungen und ausgewachsenen P. guajava-Pflanzen mit C. cubensis bestätigten dessen Pathogenität.

### Literature

- ALFENAS, A. C.; HODGES, C. S.; JENG, R., 1984: Similarities in physiological characters between Endothia eugeniae and Cryphonectria cubensis, causal agents of cankers in clove and Eucalyp-
- tus, respectively. Phytopathology 74, 841 (Abst.). BOERBOOM, J. H. A.; MAAS, P. W. T., 1970: Canker of *Eucalyptus grandis* and *E. saligna* in Surinam caused by *Endothia havanensis*. Turrialba 20, 94–99.
- BOLT, L. C., 1984 a: Climatic requirements for guavas. Farming in South Africa, B1: 1.

1984b: Guava cultivars. Farming in South Africa, C1:1.

BOLT, L. C.; ALBERTS, A. J., 1984: Pruning of guava trees. Farming in South Africa, G1: 1-4.

- BRUNER, S. C., 1916: A new species of *Endothia*. Mycologia 8, 239–242.
  CONRADIE, E.; SWART, W. J.; WINGFIELD, M. J., 1990: *Cryphonectria* canker of *Eucalyptus*, an important disease in plantation forestry in South Africa. South African Forestry Journal 152, 43-49.
- DAVIDSON, E. M.; TAY, F. S. C., 1983: Twig, branch and upper trunk cankers of Eucalyptus marginata. Plant Disease 67, 1285-1287.

DU PREEZ, S. J., 1986: Origin of the guava. Farming in South Africa, A1: 1.

- FLORENCE, E. J. M.; SHARMA, J. K.; MOHANAN, C., 1986: A stem canker disease of Eucalyptus caused by Cryphonectria cubensis in Kerala. Kerala Forest Research Institute Scientific Paper 66, 384-387.
- GIBSON, I. A. S., 1981: A canker disease of Eucalyptus new to Africa. FAO, Forest Genetics Resources Information 10, 23-24.
- HODGES, C. S., 1980: The taxonomy of Diaporthe cubensis. Mycologia 72, 542-548.
- 1988: Preliminary exploration for potential biological control agents for Psidium cattleianum.
- Technical Report 66. University of Hawaii at Manoa National Park Service. 32 pp. HODGES, C. S.; GEARY, T. F.; CORDELL, C. E., 1979: The occurrence of *Diaporthe cubensis* on *Eucalyptus* in Florida, Hawaii, and Puerto Rico. Plant Disease Reporter 63, 216–220.
- HODGES, C. S.; ALFENAS, A. C.; FERREIRA, F. A., 1986: The conspecificity of Cryphonectria cubensis and Endothia eugeniae. Mycologia 78, 343–350. MICALES, J. A.; STIPES, R. J., 1984: Differentiation of Endothia and Cryphonectria species by
- polyacrylamide gel electrophoresis. Phytopathology 74, 883–884 (Abst.). OLD, K. M.; MURRAY, D. I. L.; KILE, G. A.; SIMPSON, J.; MALAFANT, K., 1986: The pathology
- of fungi isolated from eucalypt cankers in south eastern Australia. Australian Forest Research 16, 21-36.
- PALMER, E.; PITMAN, N., 1973: Trees of Myrtle and Eucalyptus Family (Myrtaceae). In: Trees of Southern Africa V.III. Ed. by E. PALMER and N. PITMAN. A. A. Balkema, Cape Town. 2235 pp. SHARMA, J. K.; MOHANAN, C.; FLORENCE, E. J. M., 1985a: Disease survey in nurseries and
- plantations of forest tree species grown in Kerala. Research report 36. Kerala Forest Research Institute, India. 268 pp.
- 1985b: Occurrence of Cryphonectria canker disease of Eucalyptus in Kerala, India. Annals of Applied Biology 106, 265–276.
  WINGFIELD, M. J.; SWART, W. J.; ABEAR, B. J., 1989: First record of Cryphonectria canker of Eucalyptus in South Africa. Phytophylactica 21, 311–313.
- Authors' address: Dr. W. J. SWART, Ms. E. CONRADIE, Prof. M. J. WINGFIELD, Departments of Plant Pathology and Microbiology and Biochemistry, University of the Orange Free State, P. O. Box 339, Bloemfontein 9300, South Africa

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